

A DFT method for the study of the antioxidant action mechanism of resveratrol derivatives

Ali Benayahoum · Habiba Amira-Guebailia · Omar Houache

Received: 20 November 2012 / Accepted: 14 January 2013 / Published online: 3 February 2013
© Springer-Verlag Berlin Heidelberg 2013

Abstract Quantum-chemical calculations using DFT, have been performed to explain the molecular structure antioxidant activity relationship of resveratrol (RSV) (1) analogues: 3,4-dihydroxy-*trans*-stilbene (3,4-DHS) (2); 4,4'-dihydroxy-*trans*-stilbene (4,4'-DHS) (3); 4-hydroxy-*trans*-stilbene (4-HS) (4); 3,5-dihydroxy-*trans*-stilbene (3,5-DHS) (5); 3,3'-dimethoxy-4,4'-dihydroxy-*trans*-stilbene (3,3'-DM-4,4'-DHS) (6); 2,4-dihydroxy-*trans*-stilbene (2,4-DHS) (7) and 2,4,4'-trihydroxy-*trans*-stilbene (2,4,4'-THS) (8). It was found that all compounds studied were effective antioxidants with the exception of 3, 5-DHS. The high antioxidant activity of both 3, 3'-DM-4, 4'-DHS and 3, 4-DHS may be due to the abstraction of the two hydrogen atoms of the para and ortho-position hydroxyls respectively, to form a quinone structure. Our results revealed that the antioxidant pharmacophore of 2,4-DHS and 2,4,4'-THS, exhibiting higher antioxidant activity than resveratrol, is the 2-hydroxystilbene, rather than 4-hydroxystilbene. Experimental observations were satisfactorily explained and commented.

Keywords AIP · Antioxidant activity · DFT method · HOMO and LUMO · Spin density · *Trans*-resveratrol

A. Benayahoum · H. Amira-Guebailia (✉)
Laboratory of Applied Chemistry,
8 Mai 1945 Guelma University, Guelma 24000, Algeria
e-mail: amira_h_g@yahoo.co.uk

O. Houache
Petroleum & Chemical Engineering Department,
Sultan Qaboos University, P.O. Box 33, AlKhoud 123,
Muscat, Sultanate of Oman

Introduction

Polyphenols play an important role in the protection of living cells since they have a wide range of biological activities including anti-inflammatory and chemopreventive. Resveratrol (3, 5, 4'-trihydroxy-stilbene) is a naturally occurring phytoalexin present in grapes and many other plants. Many studies reported on the biological activities of resveratrol [1], but the most striking biological action, intensely investigated for the last decade is its antioxidant behavior in biological systems [2, 3]. Resveratrol has been reported to possess a potent antioxidant activity against the peroxidation of low-density lipoproteins (LDL) [4, 5] and liposomes [6], and to be a potent inhibitor of lipoxygenase [7]. Moreover, this powerful antioxidant has shown a broad range of biological effects such as neuroprotection, anti-inflammation, and anti-cancer [8, 9]. Resveratrol is able to attenuate neurodegeneration in animal models of Alzheimer's and Parkinson's diseases associated with the neuronal accumulation of β -amyloid and α -synuclein, respectively [10, 11].

Besides resveratrol, Murias et al. [12] studied the antioxidative features of many polyhydroxylated stilbenes.

The antioxidant activity of resveratrol is related to its hydroxyl (OH) groups which can scavenge free radicals produced in vivo [13, 14]. Indeed, Stivala et al. [15] reported that the deletion of all resveratrol hydroxyls as well as the substitution of these hydroxyls by OCH₃ decreased the antioxidant activity. When the 4'-hydroxyl of resveratrol was substituted by OCH₃, the EC₅₀ value of 2,2-diphenyl-1-picrylhydrazyl radical (DPPH), used to express the antioxidant activity, was 48.6. Moreover, the EC₅₀ value is 30.1

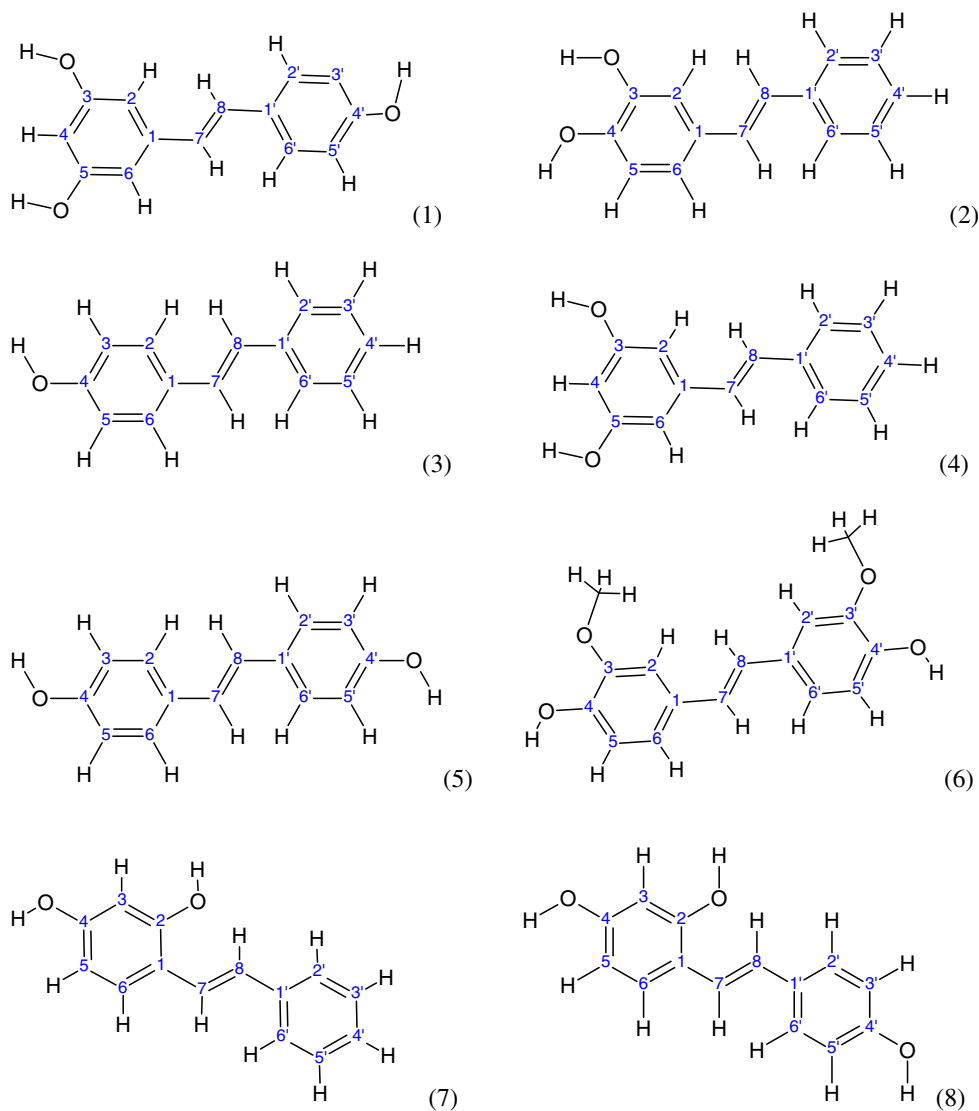
while the 3- or 5-hydroxyl is replaced by OCH_3 . These results have shown that the hydroxyl group in the 4'-position is required for the antioxidant activity, but acts synergistically with the 3 and 5-OH groups. It was also found that some resveratrol derivatives bearing orthodiphenoxyl functionality possess much higher antioxidative activity against free radical-induced peroxidation in membrane mimetic systems than those that do not possess such functionalities [14].

To find more efficient antioxidants by structural modification Cai et al. [3] synthesized resveratrol and related *trans*-stilbene analogues: 3, 4-dihydroxy-*trans*-stilbene (3, 4-DHS); 4, 4'-dihydroxy-*trans*-stilbene (4,4'-DHS); 4-hydroxy-*trans*-stilbene (4-HS) and 3,5-dihydroxy-*trans*-stilbene (3, 5-DHS) (Fig. 1) and studied in vitro, their antioxidant activity for the free radical-induced peroxidation of rat liver microsomes. They found that resveratrol and its analogues 3, 4-DHS; 4, 4'-DHS; 4-HS and 3, 5-

DHS, are effective antioxidants against both AAPH (a compound used extensively as a free radical generator and it is often used in the study of lipid peroxidation and the characterization of antioxidants) and iron-induced peroxidation of rat liver microsomes with an activity sequence of 3,4-DHS > 4,4'-DHS > resveratrol > 4-HS > 3,5-DHS. On the other hand, a kinetic analysis of the antioxidation process demonstrated that *trans*-stilbene derivatives are effective antioxidants against AAPH with the activity sequence of 3,3'-DM-4,4'-DHS > 3,4-DHS > 2,4,4'-THS > resveratrol > 3,5-DHS > 4,4'-DHS > 2,4-DHS, while the following sequence 3,4-DHS > 3,3'-DM-4,4'-DHS > 4,4'-DHS > resveratrol > 2,4-DHS > 2,4,4'-THS > 3,5-DHS was obtained for Cu^{2+} -induced low density lipoprotein peroxidation [16].

Recently, Mikulski et al. [17] studied by DFT the antioxidant activity of *trans* and *cis* resveratrol, *trans*-4,4'-dihydroxystilbene (*trans*-4,4'-DHS), *trans*-3,4-dihydroxystilbene

Fig. 1 Molecular structure of RSV (1); 3,4-DHS (2); 4-HS (3); 3,5-DHS (4); 4, 4'-DHS (5); 3,3'-DM-4,4'-DHS (6); 2,4-DHS (7) and 2,4,4'-THS (8)



(*trans*-3,4-DHS), *trans*-3,4,4'-trihydroxystilbene (*trans*-3,4,4'-THS), *trans*-3,4,5-trihydroxystilbene (*trans*-3,4,5-THS) and α , β -dihydro-3,4',5-trihydroxystilbene (α , β -dihydro-3,4',5-THS) and found that *trans*-3,4-DHS, *trans*-3,4,4'-THS, *trans*-3,4,5-THS and *trans*-4,4'-DHS exhibit higher antioxidant activity than *trans* resveratrol.

We report herein a DFT study on the antioxidative effect of resveratrol and related *trans*-stilbene analogues, 3,4-dihydroxy-*trans*-stilbene (3,4-DHS) (2); 4,4'-dihydroxy-*trans*-stilbene (4,4'-DHS) (3); 4-hydroxy-*trans*-stilbene (4-HS) (4); 3,5-dihydroxy-*trans*-stilbene (3,5-DHS) (5); 3,3'-dimethoxy-4,4'-dihydroxy-*trans*-stilbene (3,3'-DM-4,4'-DHS) (6); 2,4-dihydroxy-*trans*-stilbene (2,4-DHS) (7) and 2,4,4'-trihydroxy-*trans*-stilbene (2,4,4'-THS) (8).

Resveratrol is taken as a reference for stilbenes antioxidant activity, whereas 3,4-DHS and 4,4'-DHS, already reported to be more potent antioxidant than resveratrol by Mikulski et al. [17], are studied here, to illustrate the contribution of ortho- and para-quinone structures, generated from the original structures after H-abstraction, to the antioxidant activity. Mikulski et al. [17], for the same purpose, focused on the contribution of semiquinone structure of the phenoxy free radicals.

3,5-dihydroxy-*trans*-stilbene (3,5-DHS); 3,3'-dimethoxy-4,4'-dihydroxy-*trans*-stilbene (3,3'-DM-4,4'-DHS) (6); 2,4-

dihydroxy-*trans*-stilbene (2,4-DHS) (7) and 2,4,4'-trihydroxy-*trans*-stilbene (2,4,4'-THS) are theoretically evaluated here, for the first time, for their antioxidant action mechanism.

This study includes the determination of bond dissociation energy (BDE), adiabatic ionisation potential (AIP), the highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO), and the single electron density distribution of the radicals. It also, aims to provide a theoretical explanation of the relationship between the antioxidant activity of some stilbenes and their molecular structures. Our calculations were based on 4-HS as a pharmacophore of resveratrol antioxidant activity [18]. We further investigated the influence of the number and position of hydroxyl and methoxy groups, on the antioxidant activity of the stilbene derivatives.

Methods

All calculations were performed using GAUSSIAN 03 program package [19]. Firstly, the original structures of RSV, its analogues and the corresponding free radicals were optimized by AM1 methods [20], then all structures were fully optimized using B3LYP method at 6-31G** basis set (Fig. 2). Our analysis includes the frontier orbitals HOMO

Fig. 2 Geometries of resveratrol and its analogues ground state molecules optimized by B3LYP/6-31G**. (1) RSV; (2) 4-HS; (3) 3,5-DHS; (4) 2,4-DHS; (5) 3,4-DHS; (6) 2,4,4'-THS; (7) 4,4'-DHS and (8) 3,3'-DM4,4'-DHS

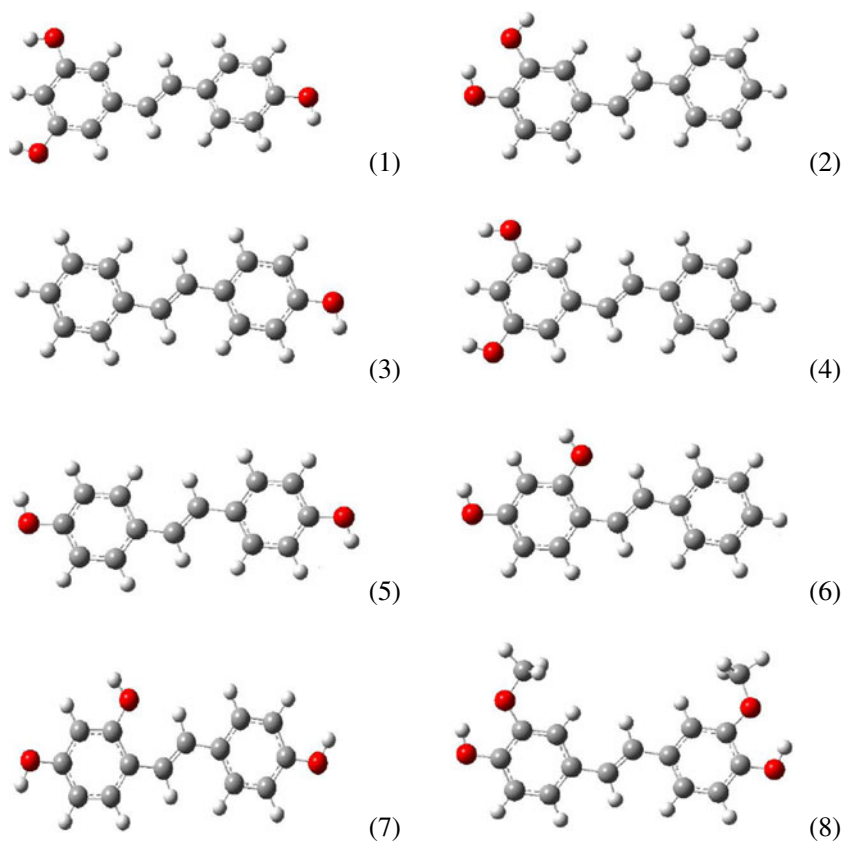


Table 1 The B3LYP/6-31G** optimized values of the dihedral angles α [°] and θ [°] and hydroxyl bond lengths [Å] of the antioxidants, studied in the gas phase

| Compounds | α | θ | 4'-OH | 3-OH | 5-OH | 4-OH | 2-OH | C=C parent | C=C 4-O-radical |
|---------------------------|----------|----------|---|---|---|------------------------------|--------|------------|-----------------|
| <i>Trans</i> -resveratrol | 0.02 | 0.02 | 0.963 ^a 0.9663 ^b | 0.962 ^a 0.9661 ^b | 0.962 ^a 0.9661 ^b | – | – | 1.349 | 1.365 |
| <i>Trans</i> -4,4'-DHS | 0.00 | 0.00 | 0.962 ^a 0.9662 | – | – | 0.962 ^a 0.9662 | – | 1.349 | 1.368 |
| <i>Trans</i> -4-HS | 0.00 | 0.00 | 0.9663 | – | – | – | – | 1.348 | 1.366 |
| <i>Trans</i> -3,4-DHS | 0.01 | 0.01 | – | 0.9662 | – | 0.9660 | – | 1.349 | 1.362 |
| <i>Trans</i> -3,5-DHS | 0.01 | 0.01 | – | 0.9658 | 0.9658 | – | – | 1.347 | 1.348 |
| <i>Trans</i> -2,4-DHS | 0.00 | 0.00 | – | – | – | 0.9661 | 0.9663 | 1.351 | 1.371 |
| <i>Trans</i> -2, 4,4'-DHS | 0.01 | 0.01 | 0.9662 | – | – | 0.9662 | 0.9663 | 1.351 | 1.372 |
| 3,3'-DM- 4,4'-DHS | 0.00 | 0.00 | 0.9699 | – | – | 0.9699 | – | 1.349 | 1.363 |

^a From ref. [17]^b From ref. [27]

and LUMO energies, AIP, BDE of phenolic hydroxyl groups on each OH site, and the spin density distribution for the radicals formed after H-removal. The conformer with the lowest electronic energy was used for calculation. Using the standard state enthalpies at 1 atm and 298.15 K, the homolytic BDE values were calculated by the following relationship:

$$\text{BDE} = H_{\text{radical}} + H_{\text{H}} - H_{\text{molecule}} \quad (1)$$

where H_{radical} is the total enthalpy of the free radical, H_{H} is the gas-phase total enthalpy of the hydrogen atom, and H_{molecule} is the total enthalpy of the parent molecule.

AIP was calculated as the total energy of the radical species after the electron oxidation minus the total energy of the species before it was oxidized:

$$\text{AIP} = E(\text{ArOH}^{\bullet+}) - E(\text{ArOH}). \quad (2)$$

Results and discussion

Molecular structure–antioxidant activity relationships of the studied stilbenes

From the optimized values of the dihedral angles α ($C_6-C_1-C_8-C_7$) and θ ($C_8-C_7-C_1-C_2$) at the B3LYP/6-31G** level (Table 1), it is evident that all resveratrol analogues studied in this work, are strictly planar. It must be pointed out that the strictly planar geometry of these compounds determines

the energetically favorable delocalization of π -electrons, and the stacking interaction of the planarity of stilbenes system may also facilitate electron transfer.

In compound 3, 4-DHS there is a weak hydrogen bond between the 3-OH and 4-O with a bond length of 2.12013 and between the 3'-OCH₃ and 4'-OH in compound 3, 3'-DM-4, 4'-DHS, with a bond length of 2.07516 Å. These H-bonds stabilize the ground state molecules and inhibit the H-abstraction reaction. However, the free radicals formed after the H-abstraction reaction in 3- and 4-positions in 3, 4-DHS include stronger intramolecular hydrogen bonds with lengths of 1.94853 and 1.9717 Å respectively, which makes the reaction occur easily. The intramolecular hydrogen bonds can be helpful for the stabilization of the 3 and 4 positions free radicals. Therefore, we can say that the higher antioxidant activity of 3, 4-DHS is due to the intramolecular hydrogen bond effect after H-abstraction.

It seems from the geometrics optimized results (Table 2), and reversible conversion from single to double bond, that all resveratrol analogues studied have a semiquinone structure after H-abstraction.

As shown in Table 2, the bond length of C_4-O in 3, 4-DHS, decreased from 1.3606 Å to 1.2530 Å, while, the bonds $C_2=C_3$ and $C_5=C_6$ decreased from 1.3842 Å and 1.3939 Å to 1.37481 Å and 1.3677 Å, respectively. On the contrary, the bonds of $C_3=C_4$, $C_4=C_5$, $C_6=C_1$ and $C_1=C_2$ increased from 1.40910, 1.39102, 1.40525, and 1.41202 to

Table 2 The B3LYP/6-31G** optimized values of the C-O, C-C and C=C bond lengths [Å] in 3, 4 DHS in the gas phase

| Compounds | C3-O | C4-O | C1-C2 | C5-C6 | C2-C3 | C3-C4 | C4-C5 | C6-C1 | C=C |
|---------------|--------|--------|--------|--------|--------|--------|--------|--------|-------|
| 3,4-DHS | 1.3772 | 1.3606 | 1.4120 | 1.3939 | 1.3842 | 1.4091 | 1.3910 | 1.4052 | 1.349 |
| 4-Semiquinone | 1.3379 | 1.2530 | 1.4369 | 1.3677 | 1.3748 | 1.4732 | 1.4423 | 1.4369 | 1.362 |
| 3,4 Quinone | 1.2232 | 1.2183 | 1.3661 | 1.3462 | 1.4599 | 1.5546 | 1.4742 | 1.4763 | 1.352 |

1.47328, 1.44239, 1.43698 and 1.43698 Å, respectively. The most stable structure for both 4 and 4'-radicals is the semiquinone form, in which the unpaired electron is disposed on the whole molecule by the double bond. The semiquinone resonance structures are favorable to stabilize the free radicals. The 5-O radicals in RSV and 3, 5- DHS, also have the same resonance structure as the 4-radical, but the unpaired electron is mainly distributed on the 5, 2, 4 and 6 atoms.

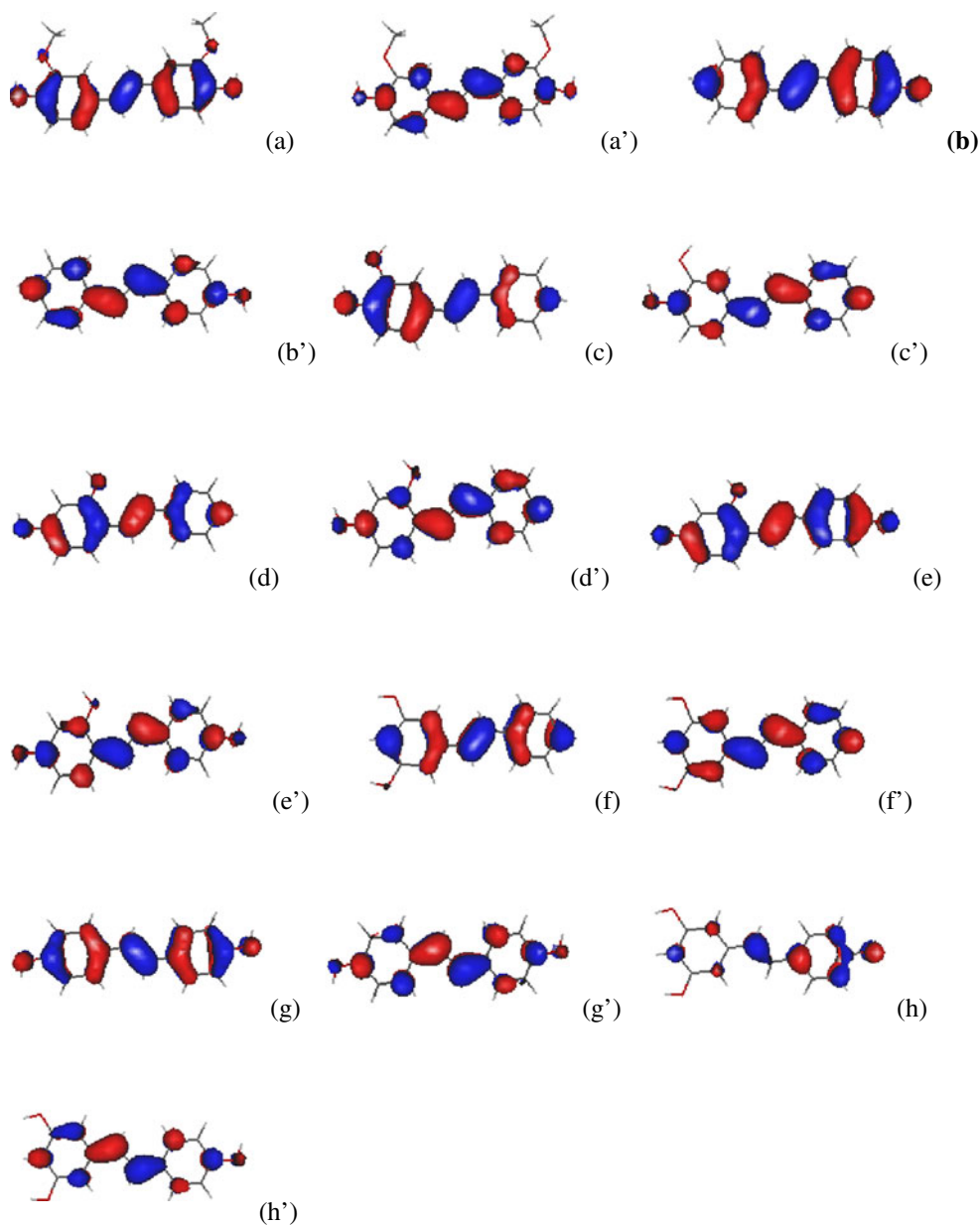
HOMO and LUMO

Both HOMO and LUMO are the main orbitals involved in the chemical reaction. The HOMO energy characterizing the

ability of electron-giving is suitable for representing the free radical scavenging potential of polyphenols, since the process to impede the auto-oxidation may involve not only the H-atom abstraction but also, the electron-transfer [21]. Higher values of ϵ_{HOMO} suggest that the molecule is a good electron donor [22]. On the other hand, the atomic sites characterized by high density of the HOMO distribution are very sensitive to the attack of free radicals and other reactive agents. The more HOMO orbital is delocalized, the more numerous are the electron sites, and more redox reactions will occur [17].

Figure 3 (a, a') shows that both HOMO and LUMO are delocalized in the whole 3,3'-DM-4,4'-DHS molecule except for C₃, C_{3'}, and C_{5'} atoms and 3, and 3'-O-

Fig. 3 The density distributions of HOMO and LUMO of 3,3'-DM-4,4'-DHS (a,a'); 4-HS (b,b'); 3,4-DHS (c, c'); 2,4-DHS (d,d'); 2,4,4'-THS (e,e'); 3,5-DHS (f,f'); 4,4' - DHS (g,g') and RSV(h,h'), respectively



methoxy groups for LUMO. Considering the disposition of HOMO and LUMO orbitals, O₄ and O_{4'} atoms of the two hydroxyls donate electrons easily so that it is possible to lose an electron during the H-abstraction reaction and form the electropositive free radical, then the proton transfer occurs. The OH groups of both 4 and 4'-positions are easily attacked by either the electrophilic or nucleophilic agents. The HOMO and LUMO electron

density (Fig. 3c) in 3, 4 DHS is mainly distributed over the molecule except for the 3-OH position for the LUMO. Regarding 4-HS and 4, 4'- DHS, the hydroxyl positions are very sensitive to the attack of free radicals and other reactive agents, due to the high density of the HOMO distribution. Noting that, the HOMO and LUMO orbitals are distributed in all hydroxyl sites, but the HOMO density is higher in 4 and 4'-OH than

Table 3 Theoretical properties of resveratrol and derivatives

| Compounds | E _{total} /Hartree | HOMO (ev) | LUMO (ev) | BDE (O-H) kJ mol ⁻¹ | AIP kJ mol ⁻¹ |
|--------------------------|-----------------------------|------------------------|------------------------|--------------------------------|--------------------------|
| 33'-DM-44'-DHS | -920.222815 | -4.8294 | -1.0003 | | 586.7466 |
| <i>Semi-quinone form</i> | -919.589188 | α=-5.0158 β=-6.0014 | α=-1.6117 β=-3.3053 | 350.1211 | |
| <i>Para-quinone form</i> | -918.967004 | -5.6071 | -3.4076 | 670.1976 | |
| 3,4-DHS | -691.170654 | -5.1698 | -1.2503 | | 632.7136 |
| <i>4-O-radical</i> | -690.550713 | α=-5.2569 β=-6.3027 | α=-1.6582 β=-3.4781 | 314.1890 | |
| <i>3-O-radical</i> | -690.544007 | α=-5.5309 β=-5.7971 | α=-1.7050 β=-3.5562 | 331.7954 | |
| <i>Quinone form</i> | -689.919739 | -6.2311 | -3.4659 | 657.3460 | |
| 4,4'-DHS | -691.169619 | -5.0112 | -1.0547 | | 613.6919 |
| <i>Semi-quinone form</i> | -690.536455 | α=-5.1606 β=-6.1707 | α=-1.4857 β=-3.3559 | 348.9055 | |
| <i>para-quinone form</i> | -689.915495 | -6.2039 | -3.7431 | 665.78047 | |
| 4, HS | -615.9493435 | -5.2420 | -1.2109 | | 640.9185 |
| <i>4-O-radical</i> | -615.314812 | α=-5.3927 β=-6.3522 | α=-1.6413 β=-3.4963 | 352.4943 | |
| 3,5-DHS | -691.168631 | -5.4972 | -0.0487 | 371.5491 | 666.76398 |
| <i>3-O-radical</i> | -690.5268428 | α=-6.0011 β=-5.8256 | α=-1.7611 β=-3.5997 | | |
| 2,4-DHS | -691.16791 | -5.0692 | -1.0187 | | 622.3615 |
| <i>4-O-radical</i> | -690.53572 | α=-5.3181 β=-6.3021 | α=-0.0541 β=-3.4661 | 346.3485 | |
| <i>2-O-radical</i> | -690.53840 | α=-5.2822 β=-5.8520 | α=-1.3896 β=-3.4577 | 339.3122 | |
| 2, 4,4'-THS | -766.38929 | -4.8509 | -0.8484 | | 596.5397 |
| <i>4-O-radical</i> | -765.75754 | α=-5.0781 β=-6.0545 | α=-1.3066 β=-3.3203 | 345.5608 | |
| <i>2-O-radical</i> | -765.760422 | α=-5.0278 β=-5.6406 | α=-1.2280 β=-3.3099 | 337.62663 | |
| <i>4'-O-radical</i> | -765.755869 | α=-4.9848 β=-6.0197 | α=-1.2751 β=-3.1946 | 349.5805 | |
| <i>para-quinone form</i> | -765.137772 | -6.1484 | -3.6596 | 658.9279 | |
| Resveratrol | -766.389138 | -5.2183 | -1.1714 | | 635.9039 |
| <i>4'-O-radical</i> | -765.755404 | α=-5.4006 β=-6.1851 | α=-1.6144 β=-3.5056 | 350.4023 | |
| <i>3-O-radical</i> | -765.745116 | α=-5.6975 β=-5.5209 | α=-1.6207 β=-3.5467 | 377.4134 | |

in 2-OH position. It was noticed that the HOMO density was absent in both 3-OH and 5-OH sites in 3, 5-DHS and RSV. If we look closely at Fig. 3, we can see that there is a significant HOMO density contribution from the double bond between C₇ and C₈ connecting the phenyl rings, in all RSV analogues. The vinyl double bond ensures π -electron delocalization between the A- and B-rings, contributes to the stabilization of RO \cdot after H-abstraction, and hence contributes to the antioxidant activity.

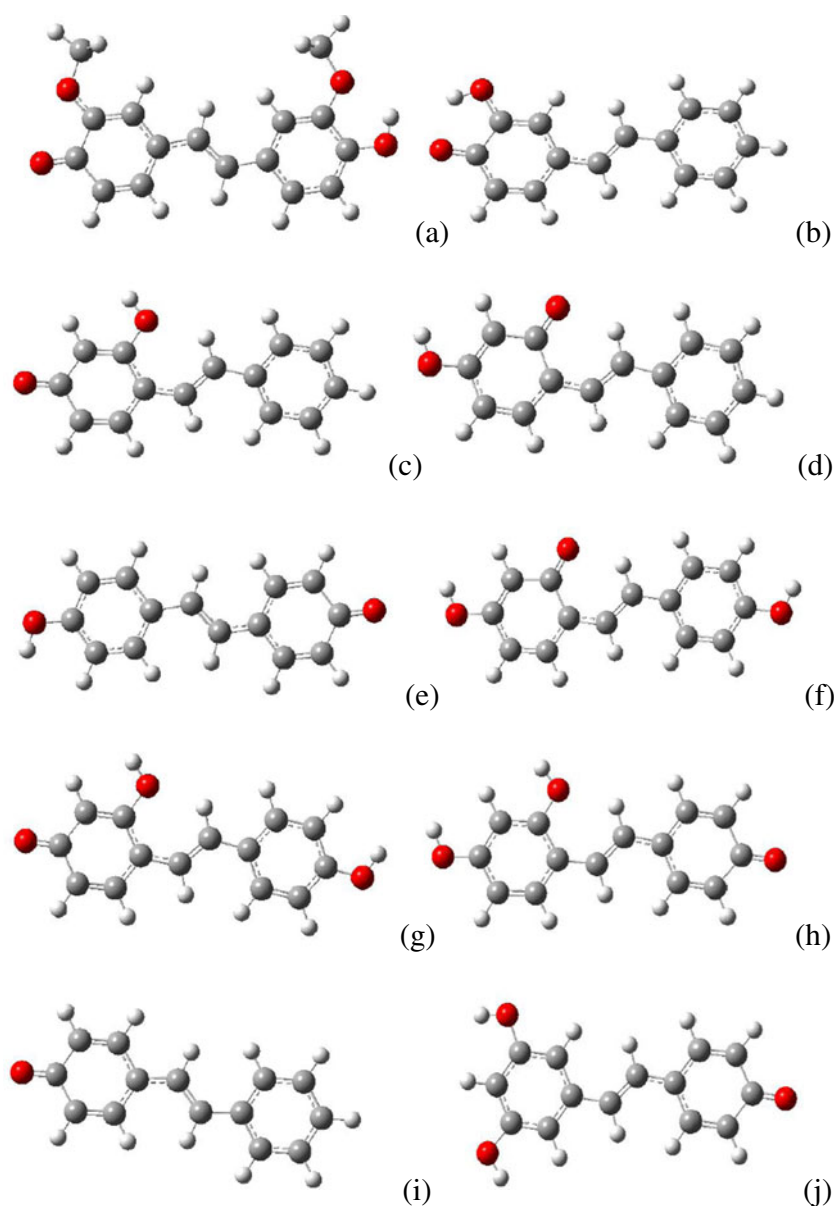
The ϵ_{HOMO} eigenvalue of 4-HS is -5.2420 eV. The substitution of a hydroxyl group in 2, 3 and 4' positions of the antioxidant pharmacophore 4-HS leads to compounds with ϵ_{HOMO} eigenvalues; -5.0692, -5.1698 and -5.0112 eV, respectively. These values are higher than that of RSV

(-5.2183 eV). The most nucleophilic compounds in this theoretical study are 3,3'-DM-44'-DHS and 2, 4, 4'-THS with ϵ_{HOMO} values of -4.8294 eV and -4.8509 eV, respectively. The results obtained in the present work indicate that the existence of a catechol structure or the addition of the methoxy groups increased HOMO values.

Adiabatic ionization potential (AIP)

Both AIP and HOMO are used to determine the electron donating ability of a molecule. The electron abstraction is the first step of the antioxidant mechanism. Therefore, molecules with a lower AIP are more active. However, low values of AIP do not guarantee high antioxidant potency of the antioxidants studied [23]. A low AIP or a high

Fig. 4 The B3LYP/6-31** fully optimized structures of the resonance forms of 4-O-radical of 3, 3'-DM4, 4'-DHS (1); 3,4-DHS (2); 2,4-DHS (3); 4, 4'-DHS (5); 2,4,4'-THS(7); 4'-O-radical of 2,4,4'-THS (8); 4-HS (9) and RSV (10); and 2-O-radical of 2,4-DHS (4) and 2,4,4'-THS(6)



HOMO reflects a strong ability to donate the electrons [24]. The results in Table 3 reveal that the highest AIP values correspond to the lowest HOMO values. For example, 3, 3'-DM-4, 4'-DHS has the highest AIP but the lowest HOMO, while 3, 5-DHS has the lowest AIP but the highest BDE.

Bond dissociation energy (BDE)

The antioxidant activity reflected in the calculated BDE values is often attributed to π -electron delocalization, leading to the stabilization of the radicals obtained after H-abstraction. This conclusion is drawn assuming that, if π electron delocalization exists in the parent molecule, it also exists in the corresponding radical [25]. In order to understand the relationship between the electron delocalization and the reactivity of the radicals, we examined the electron distribution in the singly occupied molecular orbital (SOMO), called in this case, the α -HOMO. By comparing the shape of the α -HOMO to that of the first unoccupied β orbital, we found that α -HOMO is the highest-occupied molecular orbital of spin α , and it is delocalized over the entire molecule. The shape of α -HOMO is quite similar for all radicals, and does not exhibit sufficient variations to explain the differences in activity between the OH groups. Therefore, the shape of α -HOMO is not a reliable indicator for our investigation as it does not describe the global electronic behavior of the radical.

The spin density is often considered to be a more realistic parameter and provides a better representation of the reactivity [26]. The importance of the spin density for the description of RSV antioxidant activity has been pointed out by Cao et al. [27]. We have, therefore, decided to analyze the spin density of all radicals and used Eq. (1) to calculate the hydroxyls BDEs (Table 3).

The H-abstraction reactions of hydroxyls occur easily, because of their lower BDEs (Table 3) compared to the bond energy of the covalent O-H bonds in water (498.0 kJ mol⁻¹) [28]. BDE value of 4-HS is 352.494 kJ mol⁻¹. The

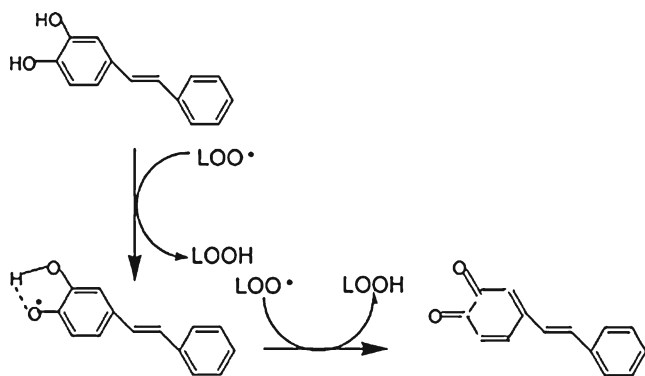


Fig. 5 Mechanism of 3, 4-DHS inhibited peroxidation

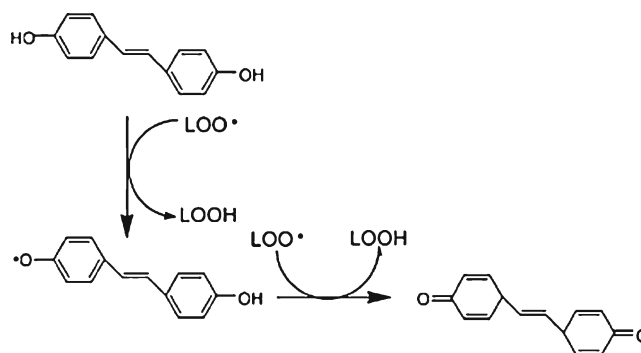


Fig. 6 Mechanism of 4, 4'-DHS inhibited peroxidation

addition of OH in 3, 2, and 4' positions of 4-HS (to obtain 3,4-DHS; 2,4-DHS; and 4,4'-DHS, respectively) decreased the 4-O-radical BDE values to 314.1890, 346.3485 and 348.9055 kJ mol⁻¹, respectively. These BDE values are lower than that of 4'-O-radical of RSV (352.1 kJ mol⁻¹). On the other hand, the BDE value for the semiquinone form of 4, 4'-DHS was 348.9055 kJ mol⁻¹, the additional OH in 2-position leading to 2,4, 4'-THS decreased BDE value of 4'-O-radical to 345.5608 kJ mol⁻¹. The addition of two methoxy groups in both 3 and 3' positions of 4, 4'-DHS increased the semiquinone BDE value to 350.1211 kJ mol⁻¹. The BDE of the 4-semiquinone form of 3,3'-DM-4,4'-DHS is higher than that of the 4, 4'DHS, despite that the spin density on the O-atom of the 4-OH radical of the 4, 4' DHS (0.3124) is higher than that in 3,3'-DM-4,4'-DHS (0.2788). This difference is related to the existence of an H-bond between the hydrogen in the 4-OH and 4'-OH groups with the oxygen atoms of the methoxy groups on C₃ and C_{3'} respectively. As a consequence, the BDEs on those sites are higher because the H-removal also implies the breaking of the H-bond. The BDE of 2-O radical in 2, 4 DHS was lower than the 4-O radical, and the BDE of 2-O radical in 2, 4, 4' THS was lower than that of 4, 4'-O radical. The unpaired electron of the 2-OH radical is on C₁ (of 2, 4 DHS and 2, 4, 4' THS), so that it can conjugate with the double bond. This indicates that the 2-O radicals are more stable than the 4, 4'-O radicals and the H-abstraction takes place easier in the 2- position than in the 4- and 4'-ones. For other stilbenes such as resveratrol, the 4'-O- radical is the most stable (Fig. 4).

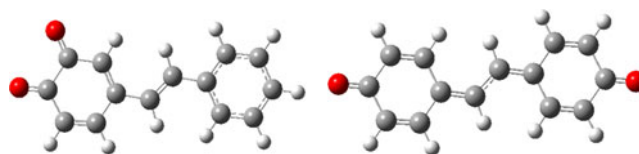
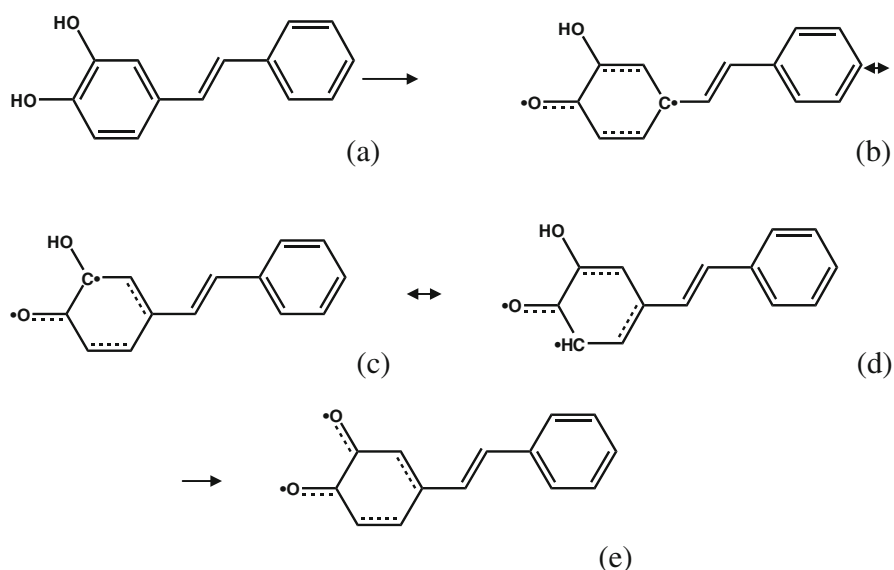


Fig. 7 The 3, 4 ortho- quinone and 4, 4'para- quinone structure of 3,4-DHS and 4, 4'-DHS

Fig. 8 The formation of semiquinone and quinone structures of 3, 4-DHS



Calculated BDE values for these compounds are in correspondence with the spin density results.

3, 4-DHS bearing ortho-diphenoxyl functionality, like rosmarinic acid [29], is remarkably more active than resveratrol and other ROHs. This is understood in view of the antioxidation mechanism of phenolic antioxidants as exemplified in Fig. 5.

As shown in Fig. 6, the 4'-OH group in 4, 4'DHS, also enhances the activity since it is able to stabilize the phenoxyl radical intermediate by resonance through the trans-double bond and be further oxidized to a para-quinone [3], nevertheless, there has been no experimental or theoretical supporting evidence.

Jovanovic et al. [30] proved experimentally that the ortho-position phenolic hydroxyls of flavonoids could undergo an abstraction of two hydrogen atoms and form a quinone structure, but there is no experimental data proving the abstraction of the second hydrogen in the para position of the 4,4'-DHS and other stilbenes, to give the para-quinone structure. We have calculated the BDE of the quinone formed by losing both hydrogen atoms in both 3, 4-ortho and 4, 4'-para-quinone positions. Figure 7 shows the geometry optimization of both ortho and para quinone structures of 3, 4-DHS and 4, 4'-DHS respectively.

We calculated the BDE of the quinone structure (Table 3) formed by losing the two hydrogen atoms in the ring A of 3,4-DHS. It is 657.3460 kJ mol⁻¹ and only 11.3615 kJ mol⁻¹ different from the BDE sum of losing 4-H and 3-H, respectively. On the other hand, the BDEs of the para 4, 4'-quinone structure of 3,3'-DM-4,4'-DHS, 4, 4'-DHS and 2,4,4'-THS were 670.1976, 665.7707 and 658.9279 kJ mol⁻¹. The half of these values are 335.0988, 332.8853 and 329.46395 kJ mol⁻¹, respectively, and are lower than those obtained for the semiquinone radicals BDEs (348.9055, 350.1211 and 345.5608 kJ mol⁻¹) (Table 3); this shows the relative stability of the para-quinone structure.

As shown in Table 2, the C₃-O and C₄-O bond lengths in 3, 4-DHS parent molecule decreased from 1.3772 and 1.3606 to 1.22322 and 1.2183 Å, in the 3, 4 quinone form.

The same change was observed in C₁-C₂ and C₅-C₆ bond lengths, decreasing from 1.4120 and 1.3939 to 1.36619 and 1.3462 Å respectively. Furthermore, C₂-C₃, C₃-C₄, C₄-C₅, and C₆-C₁, bond lengths increased from 1.3842, 1.4091, 1.3910 and 1.4052 to 1.4599, 1.5546, 1.4742 and 1.4763 Å respectively.

The HOMO and LUMO energies of the 3,4-DHS ground state molecule are -5.1698 and -1.2503 eV, respectively. After the semiquinone structure formation, the 4-O radical HOMO energy is -5.2569 eV and LUMO is -1.6582 eV, the HOMO energy of the 3-O-radical is -5.5309 eV, and the LUMO is -1.7050 eV. For the quinone structure, the HOMO and LUMO energies are -6.2311 eV and -3.4659 eV, respectively. This indicates that the semiquinone form would provide an electron again to continue the H-abstraction.

As shown in reaction mechanism (Fig. 8), the first H-abstraction in 3,4-DHS occurs in the 4-position and the radical formed will be stabilized by the semiquinone

Table 4 Spin density on the oxygen atoms in phenoxy radicals of *trans*-resveratrol and its analogues

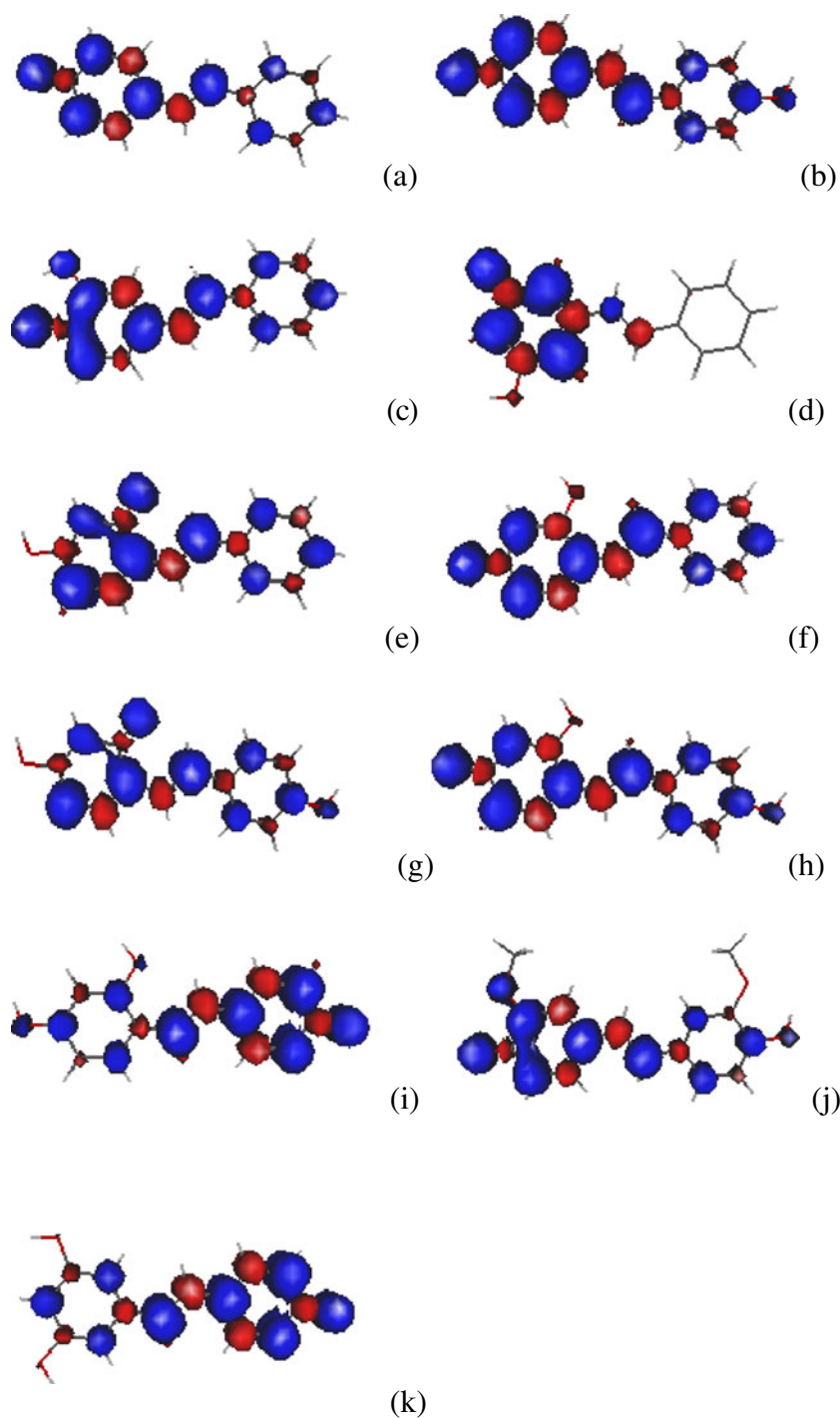
| Compounds | 4-OH | 2-OH | 3-OH | 5-OH | 4'-OH |
|---------------------------|--------|--------|--------|------|--------|
| <i>Trans</i> -resveratrol | – | – | 0.4415 | – | 0.3211 |
| 4-HS | 0.3205 | – | – | – | – |
| 3,5-DHS | – | – | 0.4143 | – | – |
| 2,4-DHS | 0.2928 | 0.2533 | – | – | – |
| 3,4-DHS | 0.2853 | – | 0.3317 | – | – |
| 2,4,4'-THS | 0.2841 | 0.2407 | – | – | 0.3119 |
| 4,4'-DHS | 0.3124 | – | – | – | – |
| 3,3'-DM-4,4'-DHS | 0.2788 | – | – | – | – |

structure (Fig. 8a and b) after passing through the three resonance structures (Fig. 8b–d). The hydrogen atom of the ortho-position hydroxyl is abstracted again and the quinone structure is formed (Fig. 8d and e).

According to the discussion above, it would be inferred that the semiquinone free radical and the 3, 4 ortho-quinone structure are the proper explanations for the higher antioxidant activity of 3, 4-DHS. The same observation was reported for resveratrol by Cao et al. [27]. Concerning the

compounds bearing 4, 4'-OH, such as 4, 4'-DHS, an H-abstraction may occur at the 4-position of the A ring and a semiquinone free radical and the 4, 4'-para quinone structure are the proper explanations for the higher antioxidant activity of 4, 4'-DHS, 3, 3'-DM4, 4'-DHS and 2, 4, 4'-THS. Moreover, the oxygen atoms of the ortho-hydroxyls (4-OH to 3-methoxy and 4'-OH to 3'-methoxy groups for 3, 3'-DM-4, 4'-DHS and 4-OH to 3-OH in 3, 4 -DHS) contribute to the single

Fig. 9 Distribution of spin densities in the radicals formed by H-removal from the A and B rings for 4-HS (1); 4, 4'-DHS (2); 3,4-DHS (3); 3,5-DHS (4); 2,4-DHS (5, 6); 2,4,4'-THS (7, 8, 9); 3, 3'-DM4, 4'-DHS (10) and resveratrol (11) computed at B3LYP/6-31G**



electron dispersing, this is why the spin density has been reduced from 0.3124 in 4, 4'-DHS to 0.2788 in 3,3'-DM-4,4'-DHS and from 0.3205 in 4-HS to 0.2853 in 3,4-DHS, respectively. The inductive effect has an important

contribution in the stabilization of the free radical as well as the hydrogen bond effect. The 2-O-radical of 2,4,4'-THS, displays a low spin density compared to the 4 and 4'-O-radicals because of the resonance effects.

Fig. 10 Spin density in the 4-O-radical of 3, 3'-DM 4, 4'-DHS (1); 3,4-DHS (2); 4, 4'-DHS (3); 4-HS (4); 2,4-DHS (5);, and 2,4,4'-THS(8); 4'-O-radical of 2,4,4'-THS (9) and resveratrol (11); 2-O-radical of 2,4-DHS (6) and 2,4,4'-DHS (7) and 3-O-radical of 3,5-DHS (10) computed at B3LYP/6-31G**

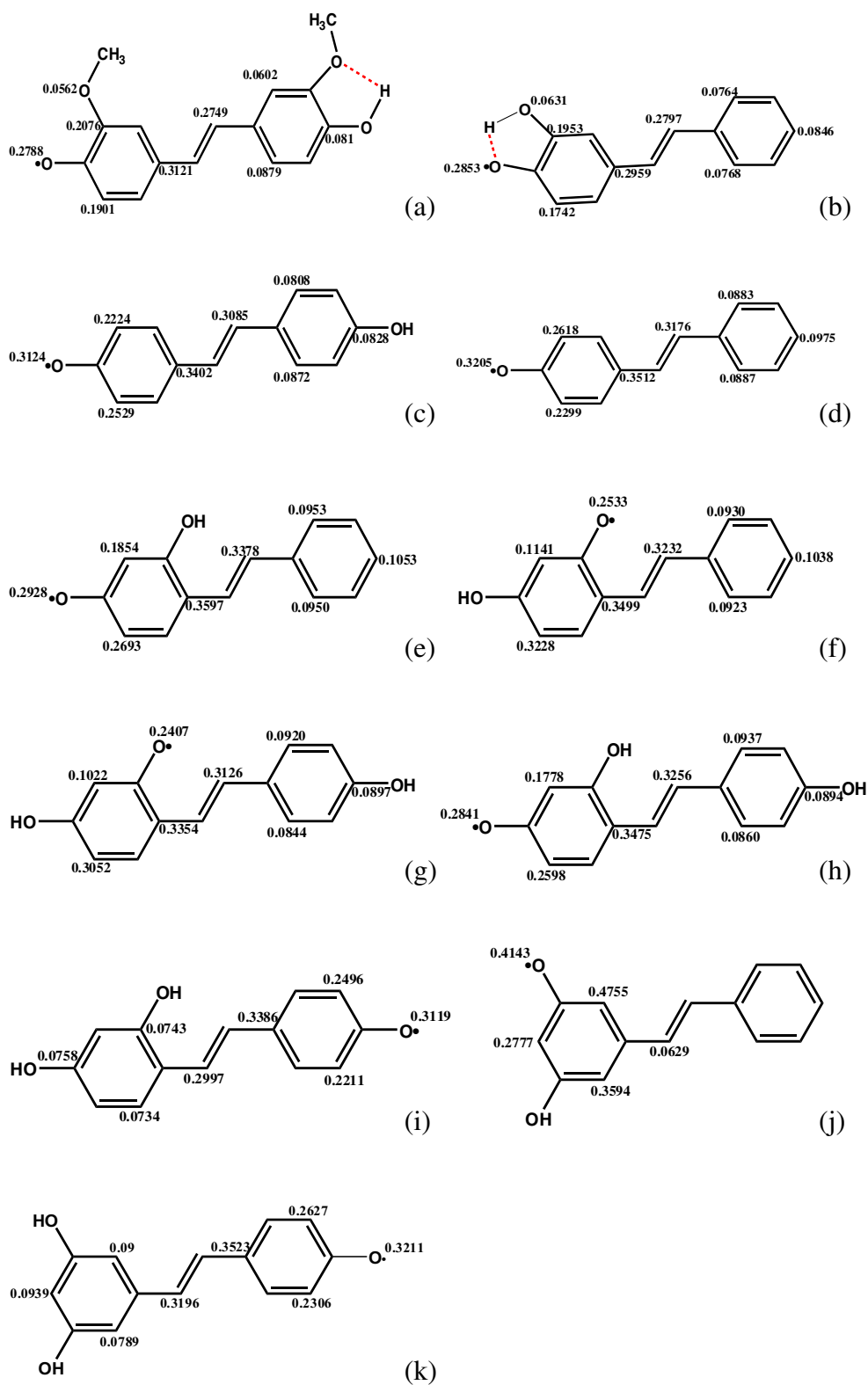
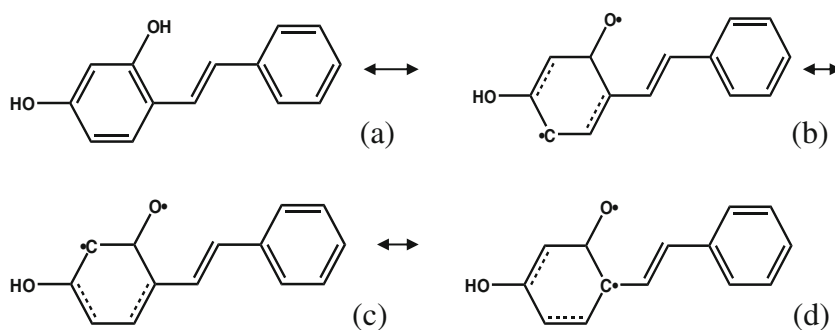


Fig. 11 The resonance structures of the 2, 4-DHS radicals



Spin density

Another molecular parameter correlated with the free radical scavenging activity is the spin density, which was also calculated. Spin density characterizes the distribution of electron spin in the free radicals and is responsible for their stability [17]. It must be stressed that the more delocalized the spin density in the radical, the easier the radical formed and thus the lower the BDE [31]. The spin density on the 4-O-radical in the 3, 4-DHS is 0.2853 (Table 4), whereas it is 0.3317 for the 3-O-radical (Fig. 9). This is a consequence of delocalization effects due to the presence of the double bond between C₇ and C₈, which allows for spin presence on the C₈. Indeed, the dominant structures of the 4-O-radicals are those in which the unpaired electron is disposed on the whole molecule. The 3-O-radical also has the same resonance structure as the 4-O-radical, but the unpaired electron is mainly distributed on the A ring (Fig. 9). Unlike the unpaired electron on C₁ of the 4-O-radical, the unpaired electron on the C₂ of the 3-O-radical can not conjugate with the double bond.

Here, it must be stressed that a hydroxyl group substituted on a carbon with positive spin density increases the stability of a radical and has the opposite effect when substituted on a carbon with negative spin density [32].

Note that the spin density on 4-O- atom in 4- HS is 0.3205 and it is delocalized through the whole molecule as shown in Fig. 9. C₁ (0.3512), C₃ (0.2618), C₅ (0.2299) and C₈ (0.3176) atoms are positive spin density centers, while C₂, C₄, C₆ are negative ones. With the addition of an OH-group on the 3-ortho position of 4- HS, the spin density on 4-O-radical decreases to 0.2853, with the delocalization on C₁ (0.2959), C₃ (0.1953), C₅ (0.1742) and C₈ (0.2797). Moreover, the 3-OH added is substituted on a carbon with positive spin density as shown in Fig. 10, so it increases the stability of the 4-O-radical. The addition of an OH-group on the 4'-para position in 4-HS, decreases the spin densities on the 4-O-radical to 0.3402 (C₁), 0.2224 (C₃), 0.2529 (C₅) and 0.3085 (C₈). In 4, 4'-DHS, 4'-OH is substituted on a carbon with positive spin density which contributes to the stability of the radical. On the other hand, 3,3'-DM-4,4'-DHS is the result of adding two methoxy groups in 4,4'-DHS on 3 and 3' -positions, the spin density on 4-O-radical decreases to

0.2788, with delocalization on C₁(0.3121), C₃ (0.2076), C₅(0.1901) and C₈ (0.2749). In view of BDEs, AIP and spin density values (Tables 3 and 4), it appears that O₃ and O₅ have almost no contribution to resveratrol antioxidant activity. As a consequence, 3, 5-DHS is not a good antioxidant.

The spin density on 4-O-radical of 2,4-DHS is 0.2928 and localized on C₁ (0.3597), C₃ (0.1854), C₅ (0.2693) and C₈ (0.3378). The addition of an OH group to the 2-position of 4-HS, unlike the other analogues, increases the spin density on C₈. It is noticed that the concentration in B-ring is slightly increased. The same finding was obtained with 2, 4, 4'-THS (addition of an OH to 4, 4'-DHS on the 2-position). On the other hand, the spin density value of 2-O-radical is 0.2533 in 2,4-DHS and 0.2407 in 2, 4, 4'-THS respectively. The later spin density values are lower than that of 3,4-DHS 4-O radical (0.2853), despite higher BDE values; 339 and 337 kJ mol⁻¹ for 2,4-DHS and 2, 4, 4'-THS respectively, compared with that of 3,4- DHS (314.1890 kJ mol⁻¹). This is the effect of the vicinity of the double bond to the 2-position in both 2, 4-DHS and 2, 4, 4'-THS. The most stable structure for these molecules is that in which the unpaired electron is localized on the ortho position of the 2-O-radical (Figs. 11 and 4). The calculated BDEs of these compounds confirm this result.

The low values of spin density in O-radicals studied show that these compounds are excellent antioxidants.

The experimental observations about some resveratrol derivatives antioxidant activity were satisfactorily explained by the results obtained by Cheng et al. [16] and Cai et al. [3]. Our results confirmed and provided deeper understanding of these experimental observations. Furthermore, we must point out that our theoretical study supports the experimental one about the 2,4,4'-THS antioxidant activity found to be higher than that of resveratrol for AAPH test but not for the Cu²⁺-induced LDL peroxidation [16].

Conclusions

The results of this DFT computations revealed that *trans*-3, 4-DHS; *trans*-3, 3'-DM4, 4'-DHS and 4, 4'-DHS exhibit

remarkably high antioxidant activity. Moreover, the later mentioned compounds are more active antioxidants than *trans*-resveratrol. However, 3,5-DHS and 4-HS are less able to scavenge free radicals than *trans*-resveratrol. These results are in full accordance with experimental results indicating that molecules bearing ortho-dihydroxyl or 4-hydroxy-3-methoxyl groups possess significantly higher antioxidant activity than those bearing no such functionalities. Our calculations demonstrate that the antioxidant potency depends on: (i) the geometry of the neutral compounds and their phenoxy radicals, (ii) the number and position of the hydroxyl groups, (iii) the semiquinone and (iv) quinone structures. It is worth noting that the inductive effect of both hydroxyl and methoxy groups, and the intramolecular hydrogen bond effect after H-abstraction contribute strongly to the antioxidant activity. The OH addition in the 2-position of 4-HS, unlike the other analogues, increases the spin density on C₈ at the double bond. On the other hand, the electron or hydrogen abstraction in ortho position is more favored than in para position for 2, 4-DHS and 2,4,4'-THS. The low spin density in the 2-position compared with the 4-position, revealed that the antioxidant pharmacophore of these two compounds is 2-hydroxystilbene.

AIP values were significantly higher than those of BDE for all compounds studied. In conclusion, this work demonstrates that H-atom transfer takes the main part in free radicals scavenging activity of the antioxidants studied and this mechanism dominates over the single-electron transfer.

Finally, this work contributes to the understanding of the pharmacological activity of the compounds studied. These findings could be useful to both food and pharmaceutical industries.

References

- Fremont L (2000) Biological effects of resveratrol. *Life Sci* 66:663–673
- Surh YJ, Hurh YJ, Kang JY, Lee E, Kong G, Lee SJ (1999) Resveratrol, an antioxidant present in red wine, induces apoptosis in human promyelocytic leukemia (HL-60) cells. *Cancer Lett* 140:1–10
- Cai YJ, Fang JG, Ma LP, Yang L, Liu ZL (2003) Inhibition of free radical induced peroxidation of rat liver microsomes by resveratrol and its analogues. *Biochim Biophys Acta (BBA)* 1637:31–38
- Tadolini B, Juliano C, Piu L, Franconi F, Cabrini L (2000) Resveratrol inhibition of lipid peroxidation. *Free Radic Res* 33:105–114
- Sánchez-Moreno C, Jiménez-Escrig A, Saura-Calixto F (2000) Study of low-density lipoprotein oxidizability indexes to measure the antioxidant activity of dietary polyphenols. *Nutr Res* 20:941–953
- Seybert DW, Milnar CM (1998) Antioxidant activity of resveratrol analogs in phospholipid oxidation. *Free Radical Bio Med* 25:S37
- Pinto MC, Garcia-Barrado JA, Macias P (1999) Resveratrol is a potent inhibitor of the dioxygenase activity of lipoxygenase. *J Agric Food Chem* 47:4842–4846
- Richard T, Pawlus AD, Iglesias ML, Pedrot E, Waffo-Teguo P, Merillon JM, Monti JP (2011) Neuroprotective properties of resveratrol and derivatives. *Ann N Y Acad Sci* 1215:103–108
- Tili E, Michaille JJ (2011) Resveratrol, microRNAs, inflammation, and cancer. *J Nucleic Acids*. doi:10.4061/2011/102431
- Vingtdeux V, Giliberto L, Zhao H, Chandakkar P, Wu Q, Simon JE, Janle EM, Lobo J, Ferruzzi MG, Davies P, Marambaud P (2010) AMP-activated protein kinase signaling activation by resveratrol modulates amyloid-beta peptide metabolism. *J Biol Chem* 285:9100–9113
- Wu Y, Li X, Zhu JX, Xie W, Le W, Fan Z, Jankovic J, Pan T (2011) Resveratrol activated AMPK/SIRT1/autophagy in cellular models of Parkinson's disease. *Neurosignals* 19:163–174
- Murias M, Handler N, Erker T, Pleban K, Ecker G, Saiko P, Szekeres T, Jager W (2004) Resveratrol analogues as selective cyclooxygenase-2 inhibitors: synthesis and structure–activity relationship. *Bioorg Med Chem* 12:5571–5578
- Wang MF, Li JG, Rangarajan M, Shao Y, Lavoie EJ, Huang TC, Ho CT (1998) Antioxidative phenolic compounds from Sage (*Salvia officinalis*). *J Agric Food Chem* 46:4869–4873
- Fang JG, Lu M, Chen ZH, Zhu HH, Li Y, Yang L, Wu LM, Liu ZL (2002) Antioxidant effects of resveratrol and its analogues against the free-radical-induced peroxidation of linoleic acid in micelles. *Chem Eur J* 8:4191–4198
- Stivala LA, Savio M, Fedarico C, Perucca P, Bianchi L, Magas G, Forti L, Pagnoni UM, Albini A, Prosperi E, Vannini V (2001) Specific structural determinants are responsible for the antioxidant activity and the cell cycle effects of resveratrol. *J Biol Chem* 276:22586–22594
- Cheng JC, Fang JG, Chen WF, Zhou B, Yang L (2006) Structure–activity relationship studies of resveratrol and its analogues by the reaction kinetics of low density lipoprotein peroxidation. *Bioorg Chem* 34:142–157
- Mikulski D, Górniak R, Molski M (2010) A theoretical study of the structure–radical scavenging activity of *trans* resveratrol analogues and *cis* resveratrol in gas phase and water environment. *Eur J Med Chem* 45:1015–1027
- Queiroz AN, Gomes BA, Moraes WM Jr, Borges RS (2009) A theoretical antioxidant pharmacophore for resveratrol. *Eur J Med Chem* 44:1644–1649
- Frisch MJ, Trucks GW, Schlegel HB et al. (2003) Gaussian 03. Gaussian, Inc., Pittsburgh
- Dewar MJS, Zoebisch EG, Healy EF, Stewart JJP (1985) AM1: a new general purpose quantum mechanical molecular model. *J Am Chem Soc* 107:3902–3909
- Nagaoka SI, Kuranaka A, Tsuboi H, Nagashima U, Mukai K (1992) Mechanism of antioxidant reaction of vitamin E: charge transfer and tunneling effect in proton-transfer reaction. *J Phys Chem* 96:2754–2761
- Fukui K (1982) Role of frontier orbitals in chemical reactions. *Science* 218:747–754
- Mikulski D, Molski M (2012) A quantum chemical study on the antioxidant activity of bioactive polyphenols from peanut (*Arachis hypogaea*) and the major metabolites of *trans* resveratrol. *Theochem* 981:38–46
- Chen W, Guo P, Song J, Cao W, Bian J (2006) The Ortho hydroxy-amino group: another choice for synthesizing novel antioxidants. *Bioorg Med Chem Lett* 16:3582–3585
- Trouillas P, Marsal P, Siri D, Lazzaroni R, Duroux JL (2006) A DFT study of the reactivity of OH groups in quercetin and taxifolin antioxidants: the specificity of the 3-OH site. *Food Chem* 97:679–688
- Szabo A, Ostlund NS (1982) Modern quantum chemistry: introduction to advanced electronic structure theory. Dover, New-York
- Cao H, Pan X, Li C, Zhou C, Deng F, Li T (2003) Density functional theory calculations for resveratrol. *Bioorg Med Chem Lett* 13:1869–1871

28. Berkowitz J, Ellison GB, Gutman D (1994) Three methods to measure RH bond energies. *Phys Chem* 98:2744–2765
29. Cao H, Cheng WX, Li C, Pan XL, Xie XG, Li TH (2005) DFT study on the antioxidant activity of rosmarinic acid. *THEOCHEM* 719:177–183
30. Jovanovic SV, Steenken S, Tosic M, Marjanovic B, Simic MJ (1994) Flavonoids as antioxidants. *J Am Chem Soc* 116:4846–4851
31. Parkinson CJ, Mayer PM, Radom L (1999) Assessment of theoretical procedures for the calculation of reliable radical stabilization energies. *J Chem Soc Perkin Trans 2*:2305–2313
32. Senthilkumar K, Kumaresan R (2012) A DFT study on the structural, electronic properties and radical scavenging mechanisms of calycosin, glycitein, pratensein and prunetin. *Theochem* 985:14–22